Model Construction and Features of Alphafold2

Lecture topics

- Input
- Evoformer module
- Structure module
- Output formatting and recycling
- Training regime and data
- Inference



Structure

Network overview



Input

• How is 3D structure encoded within the sequence?



Nicoludis, J. M., & Gaudet, R. (2018). Applications of sequence coevolution in membrane protein biochemistry. Biochimica et Biophysica Acta (BBA) - Biomembranes, 1860(4), 895–908. doi:10.1016/j.bbamem.2017.10.004

Input

• How is 3D structure encoded within the sequence?





Self-attention gives context clues to importance across the sequence, as well as the relation between difference sequences.

Pair representation to notice contextrelated clues, such as coevolution to gauge structural closeness

 Problem: How do you make a computer understand the contact network from a sequence?

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- STR**E**VKLR
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- Walk by the river bank vs.
- Get cash from the bank

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- Language models and suitability
- Self-attention networks
- Embedding space

Walk by the river bank [000010000] One-hot encoding $W_V * o. h. e$

[0.20.30.40.10.80.7] V₀: First embedded word ("walk")

Attention(V) =
$$\frac{1}{\sqrt{\dim(V)}} softmax(V * V^T) * V$$



- Language models and suitability
- Self-attention networks
- Query, Key and Value (variable embedding)





Structure module



Structure module



- 3D representations iteratively built from the evoformer pair representations with the single representation as input.
- "Rapidly develop[s] and refine[s] a highly accurate protein structure with precise atomic details. "
- Breaking the chain structure (forbidden in previous methods) and putting substantial weights on pairs from evoformer (requires evoformer to provide all information)
- Iterative refinement using recycling

Structure module

- Invariant Point Attention (IPA)
- The invariance comes from that the global transformation cancels out in the affinity computation, since L2-norm of a vector is invariant under rigid transformations.

One-hot
encoding
$$\begin{pmatrix} 1 & \cdots & 0 \\ \vdots & \ddots & \vdots \\ 0 & \cdots & 1 \end{pmatrix}$$
 \longrightarrow $W_q * o. h. e = Q$
 $W_K * o. h. e = K$
 $W_V * o. h. e = V$
Attention(Q,K,T) = $\frac{1}{\sqrt{\dim(Q,K,T)}}$ softmax $\left(Q * K^T + b - \frac{gw_c}{2} * Translation * (Q - K)\right) * V$



Recycling

- Efficiency
- Convergence properties



Training regime and data preprocessing

- For evaluation on recent PDB sequences (Figs. 2a–d, 4a, 5a), we used a copy of the PDB downloaded 15 February 2021. Structures were filtered to those with a release date after 30 April 2018 (the date limit for inclusion in the training set for AlphaFold). Chains were further filtered to remove sequences that consisted of a single amino acid as well as sequences with an ambiguous chemical component at any residue position. Exact duplicates were removed, with the chain with the most resolved Cα atoms used as the representative sequence. Subsequently, structures with less than 16 resolved residues, with unknown residues or solved by NMR methods were removed. As the PDB contains many near-duplicate sequences, the chain with the highest resolution was selected from each cluster in the PDB 40% sequence clustering of the data. Furthermore, we removed all sequences for which fewer than 80 amino acids had the alpha carbon resolved and removed chains with more than 1,400 residues. The final dataset contained 10,795 protein sequences.
- The procedure for filtering the recent PDB dataset based on prior template identity was as follows. Hmmsearch was run with default parameters against a copy of the PDB SEQRES fasta downloaded 15 February 2021. Template hits were accepted if the associated structure had a release date earlier than 30 April 2018. Each residue position in a query sequence was assigned the maximum identity of any template hit covering that position. Filtering then proceeded as described in the individual figure legends, based on a combination of maximum identity and sequence coverage.
- The MSA depth analysis was based on computing the normalized number of effective sequences (N_{eff}) for each position of a query sequence. Per-residue N_{eff} values were obtained by counting the number of nongap residues in the MSA for this position and weighting the sequences using the N_{eff} scheme⁷⁶ with a threshold of 80% sequence identity measured on the region that is non-gap in either sequence.

Inference reasoning



Recycling iteration 0, block 01 Secondary structure assigned from the final prediction

Part 2: Bioinformatics and generative modelling



General properties

- Black box appproach
- Probability and generative capabilities
- Feature importance



HMMs

 $p(y_t|x_t)$ observation probability

 $p(x_t|x_{t-1})$ transition probability

$$p(X, Y) = p(x_1) \prod_{t=1}^{T-1} p(x_{t+1}|x_t) \prod_{t'=1}^{T} p(y_{t'}|x_{t'})$$



HMMs - applications

- Sequence bioinformatics
 - MSA representation as a sequence of strings
 - MSA representation as a sequence of source outputs
 - Every sequence is a result of a random walk between sources



HMMs – modelling choices

- How many sources?
- Initialization?
 - Transition matrix helps define prior
- Optimization algorithm
 - EM algorithm

HMMs for MSA generation

- HMMer
- Distributive learning
 - Batches
 - Bayesian learning



EM algorithm

• E-step:

- Evaluate the expectation value of the observations
- M-step:
 - Given the source distribution, calculate the optimum parameter space

Expectation Maximization (EM) Algorithm $\log \text{ of expectation of } P(x|z)$ Goal: $\hat{\theta} = \underset{\theta}{\operatorname{argmax}} \log \left(\sum_{\mathbf{z}} p(\mathbf{x}, \mathbf{z} \mid \theta) \right) \quad f(\mathbf{E}[X]) \ge \mathbf{E}[f(X)]$

- 1. E-step: compute $expectation of \log of P(\mathbf{x}|\mathbf{z})$ $E_{z|\mathbf{x},\theta^{(t)}} \left[\log(p(\mathbf{x},\mathbf{z} \mid \theta)) \right] = \sum_{\mathbf{z}} \log(p(\mathbf{x},\mathbf{z} \mid \theta)) p(\mathbf{z} \mid \mathbf{x}, \theta^{(t)})$
- 2. M-step: solve

$$\theta^{(t+1)} = \underset{\theta}{\operatorname{argmax}} \sum_{\mathbf{z}} \log(p(\mathbf{x}, \mathbf{z} \mid \theta)) p(\mathbf{z} \mid \mathbf{x}, \theta^{(t)})$$

Autoregressive models

• Lab introduction



Deep Generative ML models



GAN 2014





DCGAN 2016

StyleGAN3 2021

Deep Generative ML models

- A sufficiently complex neural networks can model any regression
- Training

Generator



GANs

- Neural Networks
- Training



Validation techniques

- However, as you imply, we can additionally asses the ability of the generative algorithms in modelling the underlying process that generates data. A commonly used group of metrics for this is "information theoretic scores" that derive from the idea of <u>likelihood</u> (log-likelihood). Below are some well-known information theoretic scores:
- 1- log-likelihood (LL) score
- 2- minimum description length (MDL) score
- 3- <u>minimum message length</u> (MML) score
- 4- <u>Akaike Information Criterion</u> (AIC) score
- 5- <u>Bayesian Information Criterion</u> (BIC) score
- Note that 2, 3, 4, and 5 use some complexity penalisation factor over the LL score. This is good practice to combat <u>over-fitting</u>.

Applications in Life sciences

- Sequence analysis
- Face recognition
- Data augmentation
- Sample generation